



Correspondence

***Salmonella enterica* serovars Takoradi, Tananarive and Uganda from foods in Tripura, their virulence & antimicrobial resistance**

Sir,

The presence of *Salmonella* in foods has been considered a public health issue, as the pathogen might be transmitted from the contaminated foods into the human food chain. The zoonotic pathogen *Salmonella enterica* is a leading cause of foodborne infection in humans^{1,2}. Currently, the number of outbreaks due to *S. enterica* infections is increasing mostly due to contaminated foods of animal origin³.

The species *S. enterica* is highly diverse with about 2,600 serovars⁴. Based on the infectivity, they are classified as typhoidal (TS) and non-typhoidal *Salmonella* (NTS) serovars. Control of NTS is challenging as they have multiple sources and virulence factors. Gastroenteritis caused by the NTS is self-limiting, but it also instigates significant health risks due to its invasive nature⁵. The incidence of invasive non-typhoidal *Salmonella* (iNTS) has been increasing in several countries, especially Malawi⁶⁻⁸. Globally, the infections and deaths caused by NTS have been projected to be about 78 million and >59,000, respectively⁹.

NTS control requires a 'One Health Approach', as involving multiple sectors such as public health, animal welfare, and food and environment protection remains essential. The NTS infections in India are on the rise¹⁰. Several virulence factors in TS and NTS encoded on the *Salmonella* pathogenicity islands (SPIs) play an important role in invasion into the host cell, multiplication and spread¹¹.

Since 2020, the Indian Council of Medical Research (ICMR) has conducted an ongoing project on systematic laboratory-based surveillance of foodborne enteric disease and related outbreaks in four North-east Indian States. Between January and April 2023, we screened 1004 food samples collected from 36 places covering 4 different regions of Tripura. During the surveillance period, there was no flux of foreign

travellers. In this ongoing study, we identified three *S. enterica* serovars that were not reported from India. These *Salmonella* strains were first identified by biochemical tests¹², followed by serotyping using *Salmonella* somatic (O) and flagellar (H) antisera (SSI Diagnostica, Denmark) according to the Kauffmann–White scheme⁴. Antimicrobial susceptibility was determined by disk diffusion testing¹³. To determine the important virulence encoding genes, these strains were subjected to PCR analysis following previously published methods^{14,15}.

S. enterica serovars Takoradi, Tananarive and Uganda were identified from market samples of mutton, sweet and fish, respectively, collected between January and April 2023 in Tripura. The serotype scheme, antimicrobial resistance profile and virulence-encoding genes detected by the PCR assay are shown in Table. *S. Takoradi* and *S. Tananarive* were resistant to tetracycline. All these serovars showed reduced susceptibility to azithromycin but were susceptible to fluoroquinolones (ciprofloxacin, norfloxacin, ofloxacin), meropenem, ceftriaxone, sulfamethoxazole/trimethoprim, chloramphenicol and cefotaxime. The emergence of reduced susceptibility to azithromycin is a cause of concern. This drug has been used to treat enteric infections in India. Many of the TS and NTS clinical strains have shown resistance to azithromycin^{16,17}.

All the three serovars harboured *orgA*, (oxygen-regulated gene for host recognition and invasion), *invE/A* (SPI-1, for host cell invasion), *trrC* (tetrathionate reductase, fitness advantage), *ssaQ* (SPI-2 T3SS; secretion system apparatus protein), *mgtC* (SPI-3 intracellular proliferation inside macrophages, Mg²⁺ transporter), *misL* (SPI-3, T5SS-secreted (autotransported) adhesins, involved in intramacrophage survival), *spi4R* and *spi4D* (encodes a T1SS), *sopB* (SPI-1, SPI-5-T3SS secreted effector *Salmonella* outer protein B for bacterial internalization),

Table. Detection of antimicrobial resistance and virulence encoding genes in *S. Takorandi*, *S. Tananarive* and *S. Uganda*

Sample ID	Source	<i>Salmonella enterica</i> serovar (O:H antigen formula)	Antimicrobial susceptibility test			Virulence encoding gene*
			Resistance	Intermediate	Susceptible	
TRP/NTH/ RAWMEAT/1829	Mutton	Takoradi (8:i;1,5)	TET	AM, S, AZM, D, CAZ	NA, CIP, NOR, OFX, MEM, CR, SXT, C, CTX	<i>invE/A, orgA, ttrC, ssaQ, mgtC, misL, spi4R, spi4D, sopB, pipA, hilA, stn</i>
TRP/STH/ SWEET/1966	Sweet	Tananarive (6,8:y;1,5)	TET	AM, S, AZM, D	NA, CIP, NOR, OFX, MEM, CR, SXT, C, CAZ, CTX	<i>invE/A, orgA, ttrC, ssaQ, mgtC, misL, spi4R, spi4D, sopB, pipA, hilA, stn</i>
TRP/DHL/ RAWFISH/1104	Fish	Uganda (3,10:l,z ₁₃ ;1,5)		AM, S, AZM, D, NA, TET	CIP, NOR, OFX, MEM, CRO, SXT, C, CAZ, CTX	<i>invE/A, orgA, ttrC, ssaQ, mgtC, misL, spi4R, spi4D, sopB, pipA, pefA, hilA, stn</i>

*All the serovars were negative for *spvC*. AM, ampicillin; AZM, azithromycin; C, chloramphenicol; CAZ, ceftazidime; CIP, ciprofloxacin; CRO, ceftriaxone; CTX, cefotaxime; D, doxycycline; MEM, meropenem; NA, nalidixic acid; NOR, norfloxacin; OFX, ofloxacin; S, streptomycin; SXT, sulfamethoxazole/trimethoprim; TET, tetracycline

pip (SPI-5, pathogenicity island protein for intestinal mucosal fluid secretion and inflammation), *hilA* (SPI-1-T3SS, hyperinvasive locus the expression of invasion) and *stn* (*Salmonella* enterotoxin). In addition to these potential virulence genes, the *S. Uganda* serovar had *pefA* that encodes fimbria-associated virulence. The possession of these virulence encoding genes would suggest that the NTS serovars investigated in the current study have the ability to cause enteric salmonellosis in humans. Many of these genes have been reported to be present in *S. Typhi*. In this study, all the serovars were negative for the plasmid virulence factor encoding gene *spvC*. Some of the studies indicate importance of this gene for the survival and proliferation of NTS inside the reticuloendothelial cells and associated with bacteraemia in humans^{18,19}.

To our knowledge, *S. enterica* serovars Takoradi, Tananarive and Uganda have not yet been reported from India. *S. Takoradi* has been reported from poultry slaughterhouses in Korea²⁰, and birds in Scotland²¹. *S. Tananarive* has been isolated from humans²² and food animals (<https://www.fao.org/4/i1547e/i1547e00.pdf>, <https://www.bvmj.bu.edu.eg/issues/25-2/8.pdf>). *S. Uganda* caused a foodborne outbreak in China²³ and was also identified from turkeys in Canada²⁴, cattle and mink in USA^{25,26}, camels in Nigeria²⁷, and ducks in Trinidad²⁸.

Our findings highlight the risks associated with new *Salmonella* serovars with virulence potential and antimicrobial resistance in foods. Considering the public health risks associated with food contamination, constant surveillance and stringent control measures

are needed at the production level and also along the food chain.

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References

- Majowicz SE, Musto J, Scallan E, Angulo FJ, Kirk M, O'Brien SJ, *et al.* The global burden of non-typhoidal *Salmonella* gastroenteritis. *Clin Infect Dis* 2010; 50 : 882-89.
- Kirk MD, Pires SM, Black RE, Caipo M, Crump JA., Devleeschauwer B, *et al.* World Health Organization estimates of the global and regional disease burden of 22 foodborne bacterial, protozoal, and viral diseases, 2010: A data synthesis. *PLoS Med* 2015; 12 : e1001921.
- European Food Safety Authority (EFSA). Scientific Opinion on the risk posed by pathogens in food of non-animal origin. Part 2 (*Salmonella* and Norovirus in leafy greens eaten raw as salads). *EFSA J* 2014; 12 : 3600.
- Institut Pasteur. World Health Organization Collaborating Centre for Reference and Research on *Salmonella*. Antigenic formulae of the *Salmonella* serovars. Available from: https://www.pasteur.fr/sites/default/files/veng_0.pdf, accessed on July 26, 2023.
- Acheson, D, Hohmann, EL. Nontyphoidal salmonellosis. *Clin Infect Dis* 2001; 32 : 263-69.
- Feasey NA, Dougan G, Kingsley RA, Heyderman RS, Gordon MA. Invasive non-typhoidal *Salmonella* disease: An emerging and neglected tropical disease in Africa. *Lancet* 2012; 379 : 2489-99.
- Marchello CS, Fiorino F, Pettini E, Crump JA, Vacc-iNTS Consortium Collaborators. Incidence of non-typhoidal *Salmonella* invasive disease: A systematic review and meta-analysis. *J Infect* 2021; 83 : 523-32.
- Gordon MA, Graham SM. Invasive salmonellosis in Malawi. *J Infect Dev Ctries* 2008; 2 : 438-42.
- Havelaar AH, Kirk MD, Torgerson PR, Gibb HJ, Hald T, Lake RJ, *et al.* World Health Organization global estimates and regional comparisons of the burden of foodborne disease in 2010. *PLoS Med* 2015; 12 : E1001923.
- Dudhane RA, Bankar NJ, Shelke YP, Badge AK. The rise of non-typhoidal *Salmonella* infections in India: causes, symptoms, and prevention. *Cureus* 2023; 15 : e46699.
- Jajere SM. A review of *Salmonella enterica* with particular focus on the pathogenicity and virulence factors, host specificity and antimicrobial resistance including multidrug resistance. *Vet World* 2019; 12 : 504-21.
- Indian Council of Medical Research, Government of India. *Standard operating procedures*. Available from: https://www.icmrfoodnet.in/static/assets/files/ICMR_StandardOperatingProcedures.pdf, accessed on July 29, 2024.
- Clinical and Laboratory Standards Institute. *M100. Performance standards for antimicrobial susceptibility testing*. 33rd edition. Available from: https://clsi.org/media/tc4b1paf/m10033_samplepages-1.pdf, accessed on July 29, 2023.
- Soto SM, Rodríguez I, Rodicio MR, Vila J, Mendoza MC. Detection of virulence determinants in clinical strains of *Salmonella enterica* serovar enteritidis and mapping on macro restriction profiles. *J Med Microbiol* 2006; 55 : 365-73.
- Gharieb RM, Tartor YH, Khedr MH. Non-typhoidal *Salmonella* in poultry meat and diarrhoeic patients: Prevalence, antibiogram, virulotyping, molecular detection and sequencing of class I integrons in multidrug resistant strains. *Gut Pathog* 2015; 7 : 34.
- Jain P, Viswanathan R, Halder G, Basu S, Dutta S. Draft whole-genome sequences of two multidrug-resistant *Salmonella enterica* serovar Senftenberg sequence type 14 strains resistant to ciprofloxacin, ceftriaxone, and/or azithromycin, isolated from Kolkata, India. *Microbiol Resour Announc* 2022; 11 : e0097821.
- Carey ME, Jain R, Yousuf M, Maes M, Dyson ZA, Thu TNH, *et al.* Spontaneous emergence of azithromycin resistance in independent lineages of *Salmonella* Typhi in Northern India. *Clin Infect Dis* 2021; 72 : e120-e127.
- Libby S, Adams LG, Ficht TA, Allen C, Whitford HA, Buchmeier NA, *et al.* The *spv* Genes the *Salmonella* Dublin virulence plasmid are required for severe enteritis and systemic infection in the natural host. *Infect Immun* 1997; 65 : 1786-92.
- Gopinath A, Allen TA, Bridgwater CJ, Young CM, Worley MJ. The *Salmonella* type III effector SpvC triggers the reverse transmigration of infected cells into the bloodstream. *PLoS One* 2019; 14 : e0226126.
- Kidie DH, Bae DH, Lee YJ. Prevalence and antimicrobial resistance of *Salmonella* isolated from poultry slaughterhouses in Korea. *Jpn J Vet Res* 2013; 61: 129-36.
- Fricker CR. A note on *Salmonella* excretion in the black headed gull (*Larus ribibundus*) feeding at sewage treatment works. *J Appl Bacteriol* 1984; 56 : 499-502.
- Phuong TLT, Rattanavong S, Vongsouvath M, Davong V, Lan NPH, Campbell JI, *et al.* Non-typhoidal *Salmonella* serovars associated with invasive and non-invasive disease in the Lao People's Democratic Republic. *Trans R Soc Trop Med Hyg* 2017; 111 : 418-24.
- Yan AX, Kang Y, Cui Y, Zhao WX, Li SF, Wang M, *et al.* Etiological analysis on a foodborne disease outbreak caused by two serotypes of *Salmonella*. *Zhonghua Liu Xing Bing Xue Za Zhi* 2023; 44 : 1440-46.
- Sodagari HR, Shrestha RD, Agunos A, Gow SP, Varga C. Comparison of antimicrobial resistance among *Salmonella enterica* serovars isolated from Canadian turkey flocks, 2013 to 2021. *Poult Sci* 2023; 102 : 102655.
- Brichta-Harhay DM, Arthur TM, Bosilevac JM, Kalchayanand N, Shackelford SD, Wheeler TL, *et al.* Diversity of multidrug-resistant *Salmonella enterica* strains associated with cattle at harvest in the United States. *Appl Environ Microbiol* 2011; 77: 1783-96.
- Agga GE, Silva PJ, Martin RS. Prevalence, serotypes, and antimicrobial resistance of *Salmonella* from mink feces and feed in the United States. *Foodborne Pathog Dis* 2022; 19: 45-55.
- Raufu IA, Odetokun IA, Oladunni FS, Adam M, Kolapo UT, Akorede GJ, *et al.* Serotypes, antimicrobial profiles, and public health significance of *Salmonella* from camels slaughtered in Maiduguri central abattoir, Nigeria. *Vet World* 2015; 8: 1068-72.
- Rampersad J, Johnson J, Brown G, Samlal M, Ammons D. Comparison of polymerase chain reaction and bacterial culture for *Salmonella* detection in the Muscovy duck in Trinidad and Tobago. *Rev Panam Salud Publica* 2008; 23: 264-67.